

Tetrodotoxin Framework Construction from Linear Substrates Utilizing a Hg(OTf)₂-Catalyzed Cycloisomerization Reaction: Synthesis of the Unnatural Analogue 11-*nor*-6,7,8-Trideoxytetrodotoxin

Keisuke Nishikawa, Takayuki Noguchi, Seiho Kikuchi, Takahiro Maruyama, Yusuke Araki, Mari Yotsu-Yamashita, and Yoshiki Morimoto

Citation	Organic Letters. 23 (5); 1703 - 1708
Issue Date	2020-03
Type	Journal Article
Textversion	Author
Supporting Information	<p>The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.orglett.1c00125.</p> <p>Experimental procedures, the spectroscopic data, and copies of ¹H and ¹³C NMR spectra (PDF)</p>
Rights	<p>This document is the Accepted Manuscript version of a Published Work that appeared in final form in Organic Letters, copyright © American Chemical Society after peer review and technical editing by the publisher. To access the final edited and published work see https://doi.org/10.1021/acs.orglett.1c00125.</p>
DOI	10.1021/acs.orglett.1c00125

Self-Archiving by Author(s)
Placed on: Osaka City University Repository

Nishikawa, K., Noguchi, T., Kikuchi, S., Maruyama, T., Araki, Y., Yotsu-Yamashita, M., & Morimoto, Y. (2021). Tetrodotoxin Framework Construction from Linear Substrates Utilizing a Hg(OTf)₂-Catalyzed Cycloisomerization Reaction: Synthesis of the Unnatural Analogue 11-*nor*-6,7,8-Trideoxytetrodotoxin. *Organic Letters*, 23(5), 1703–1708. <https://doi.org/10.1021/acs.orglett.1c00125>

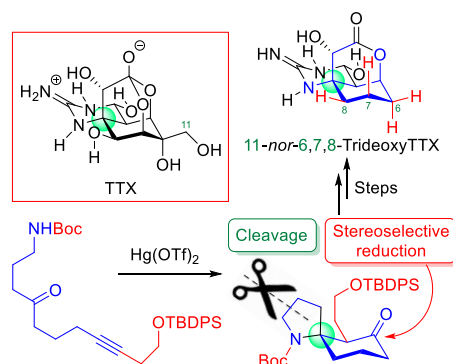
Tetrodotoxin Framework Construction from Linear Substrates Utilizing a $\text{Hg}(\text{OTf})_2$ -Catalyzed Cycloisomerization Reaction: Synthesis of Unnatural Analogue 11-*nor*-6,7,8-Trideoxytetrodotoxin

Keisuke Nishikawa,^{*,†} Takayuki Noguchi,[†] Seiho Kikuchi,[†] Takahiro Maruyama,[†] Yusuke Araki,[†] Mari Yotsu-Yamashita,[‡] and Yoshiki Morimoto^{*,†}

[†]Department of Chemistry, Graduate School of Science, Osaka City University, Sumiyoshi-ku, Osaka 558-8585, Japan.

[‡]Graduate School of Agricultural Science, Tohoku University, Sendai, Miyagi 980-8572, Japan

Supporting Information Placeholder



ABSTRACT: In this contribution, we propose a new synthetic approach to tetrodotoxin (TTX), one of the most famous marine toxins, that, after first preparing a functionalized linear substrate, forms the cyclohexane core from the substrate utilizing our mercuric triflate ($\text{Hg}(\text{OTf})_2$)-catalyzed cycloisomerization reaction. The concept was applied to the synthesis of 11-*nor*-6,7,8-trideoxyTTX and 11-*nor*-4,9-anhydro-6,7,8-trideoxyTTX, unnatural TTX analogues, demonstrating the validity of our new approach.

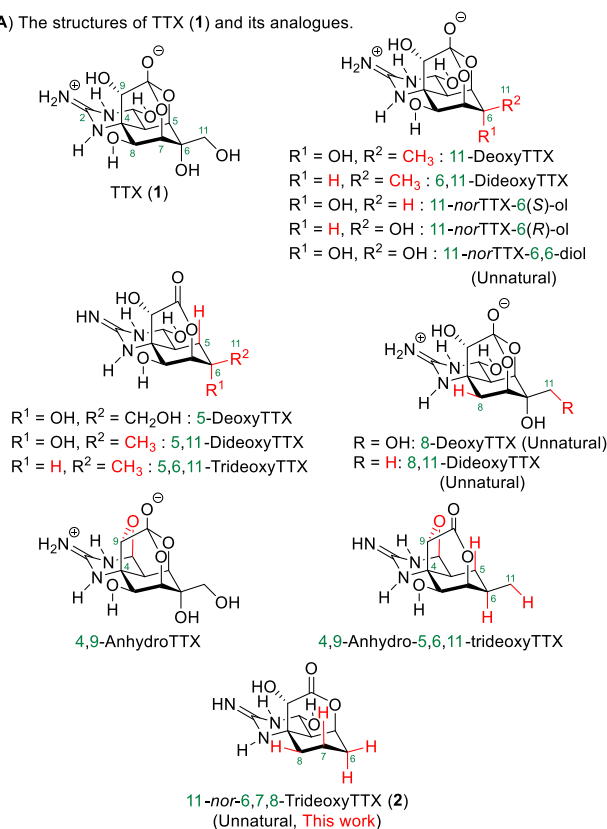
Tetrodotoxin (**1**, TTX, [Scheme 1A](#)) is one of the best-known marine natural products and was first isolated from the ovaries of globefish by Tahara in 1909.¹ Its unique and complex structure was determined by three groups,² and was subsequently established via X-ray crystallographic analysis of its derivative.³ The structural features that contain a dioxo-adamantane skeleton functionalized by a number of hydroxy groups and a cyclic guanidine in a small molecule make it the most difficult synthetic target in terms of total synthesis. TTX also exhibits a specific block of sodium ion influx through the sodium ion channel proteins.⁴ The TTX family includes various analogues, largely deoxy and anhydro types ([Scheme 1A](#)).⁵ The total synthesis of TTX and its natural analogues has been conducted several times,⁶ whereas the synthesis of its unnatural analogues has also been reported in terms of the studies on the biochemical behavior of compounds with substituent patterns unavailable in natural form. For example, Isobe and Nishikawa et al. reported the synthesis of 8,11-dideoxyTTX^{6l-n} and 8-deoxyTTX^{6q} as unnatural analogues and demonstrated the role of the hydroxy group at the C8 position in sodium channel inhibition. Therefore, establishing a

synthetic pathway for TTX analogues is crucial to elucidate the inhibition activity of voltage-gated sodium channels and the biosynthetic pathways for TTX.

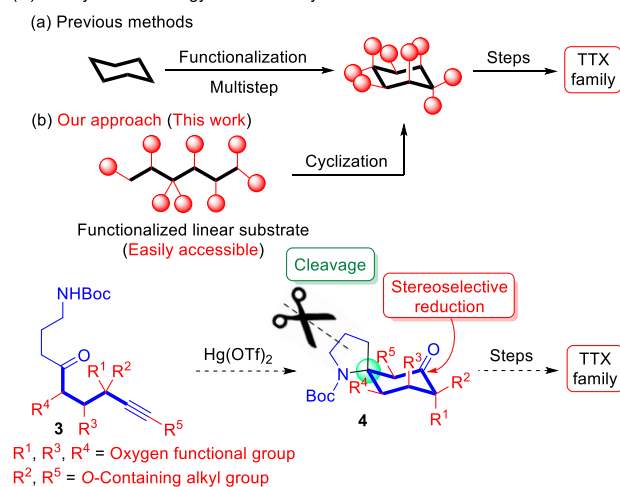
In fact, the total synthesis of TTX has been reported by five research groups.⁶ All of the synthetic strategies reported thus far have adopted the construction of the cyclohexane core at the early stage; however, the transformations of dense functional groups on the cyclohexane core resulted in numerous reaction steps due to the attendant steric hindrance ([Scheme 1B](#)). Considering this, we aimed to synthesize the TTX-type functionalized linear substrate at the early stage of synthesis to minimize the number of extra steps of transformation on the congested cyclohexane core. Following this, the mid-stage cyclization could construct the cyclohexane core and the latter conversion could facilitate the access to TTX. The key cyclization is our $\text{Hg}(\text{OTf})_2$ -catalyzed cycloisomerization reaction⁷ that constructs a 1-azaspiro[4.5]decane skeleton with a nitrogen-containing tetrasubstituted carbon from linear substrates in high stereoselectivity and good yield via a Petasis–Ferrier-type reaction. In this plan, the

Scheme 1. Structures of TTX (1) and Its Analogues, and (B) Our Synthetic Strategy for the TTX Family

(A) The structures of TTX (1) and its analogues.



(B) Our synthetic strategy for TTX family

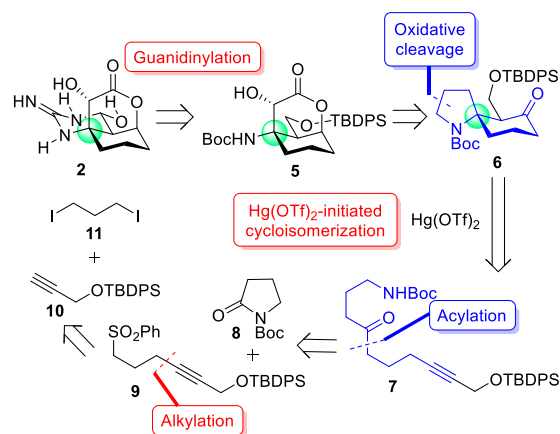


functionalized linear substrate **3** should be treated with $\text{Hg}(\text{OTf})_2$ to yield the key intermediate spirocycle **4**, which would be led to the TTX family through the stereoselective reduction of a carbonyl moiety and the carbon–carbon bond oxidative cleavage on a pyrrolidine ring followed by intramolecular cyclization to construct a lactone ring. If certain linear substrates with various oxygen functional groups could be synthesized, the synthetic strategy for the desired TTX analogues could be established by the cycloisomerization producing the corresponding spirocycles. In this article, we report the synthesis of 11-nor-6,7,8-trideoxyTTX (**2**), an unnatural analogue of TTX, as the first model compound (Scheme 1A). Since the 7-deoxy analogue has been

produced neither naturally nor through synthesis, the role of the hydroxy group at the C7 position in sodium channel inhibition remains unclear. The validity of our synthetic strategy would be confirmed by the synthesis of analogue **2**.

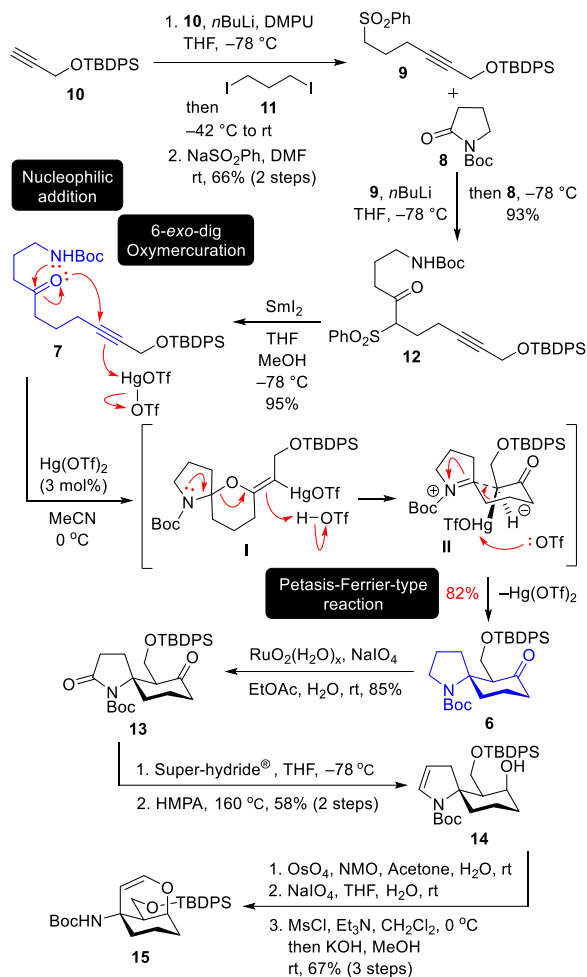
The retrosynthetic analysis of analogue **2** is shown in Scheme 2. Here target **2** would be synthesized through guanidinylation from lactone **5**, which would be derived from spirocycle **6** via the carbon–carbon bond oxidative cleavage. Compound **6** with a nitrogen-containing tetrasubstituted carbon would be then constructed by the key cycloisomerization of linear ynone **7**, which would be prepared through acylation of sulfone **9** with easily accessible pyrrolidinone **8**. We predicted that compound **9** would be synthesized by alkylation of a lithium acetylide of the known alkyne **10** with 1,3-diiodopropane (**11**).

Scheme 2. Retrosynthetic Analysis of 11-nor-6,7,8-TrideoxyTTX (2)



The synthesis commenced with the known alkyne **10**, derived from commercially available 2-propyn-1-ol (Scheme 3).⁸ Sulfone **9** was synthesized through alkylation of a lithium acetylide of **10** with commercially available 1,3-diiodopropane (**11**) and subsequent sulfonylation. Following the acylation of an α -anion of **9** with the known pyrrolidinone **8**,⁹ which can be easily prepared from commercially available 2-pyrrolidone, the cyclization precursor **7** was prepared through Sml_2 -mediated desulfonylation¹⁰ of **12**. The key $\text{Hg}(\text{OTf})_2$ -catalyzed cycloisomerization reaction⁷ of **7** produced the desired spirocycle **6** in a stereoselective manner (82%).^{11,12} The aminoketal **I** would be formed through a 6-*exo*-dig intramolecular oxymercuration to the alkyne π -electron activated by coordination of the catalyst followed by the nucleophilic addition of the nitrogen function. The intermediate **I** could be cleaved via protonation with the generated TfOH to give iminium ion intermediate **II** (Petasis–Ferrier-type cyclization). The construction of a carbocycle would provide spirocycle **6** with regeneration of the catalyst. Considering certain chair-like transition states, the desired **6** would diastereoselectively be obtained by way of the more stable transition state **II** with less steric repulsion.¹³ Following the oxidation using RuO_4 to afford amide **13**,¹⁴ the hydride reduction using Super-hydride[®] produced the desired diol, which was heated in HMPA to give enecarbamate **14** as a sole diastereomer.¹⁵ Next, we

Scheme 3. Synthesis of Enol Ether 15



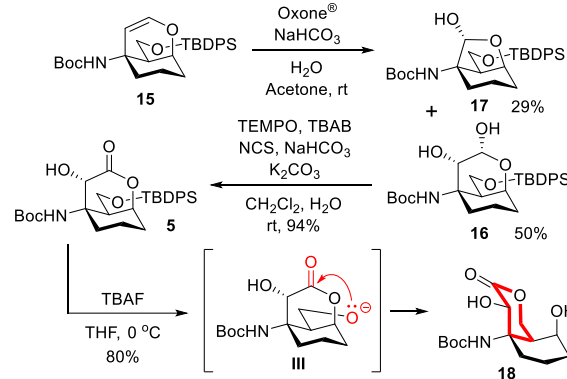
intended to cleave a double bond in **14**. After **14** was treated with OsO₄ and NMO, the oxidative cleavage of the resultant diol using NaIO₄ followed by intramolecular cyclization resulted in the desired lactol. Following dehydration using MsCl and Et₃N, the desired enol ether **15** was obtained via removal of a formyl group.

Next, we focused on the synthesis of analogue **2** from **15** (Scheme 4A). Dihydroxylation using Oxone® and NaHCO₃ produced the desired diol **16** in 50% yield, along with byproduct **17** which would oxidatively be cleaved from **16** (29%).^{16,17} The selective TEMPO oxidation¹⁸ of **16** produced lactone **5** in good yield (94%). However, when a *tert*-butyldiphenylsilyl (TBDPS) protecting group in **5** was removed using tetra-*n*-butylammonium fluoride (TBAF), only undesired lactone **18** was obtained in 80% yield due to the unexpected intramolecular nucleophilic attack. In 2015, Xu and Ciufolini reported a similar recyclization reaction in a formal synthesis of TTX (Scheme 4B).¹⁹ To prevent this unwanted nucleophilic attack, we increased the number of methylene units in the side chain when synthesizing lactone **21** (Scheme 4C). It is presumed that the recyclization would not proceed by the TBDPS deprotection in **21** because the seven-membered lactone **22** is difficult to generate.

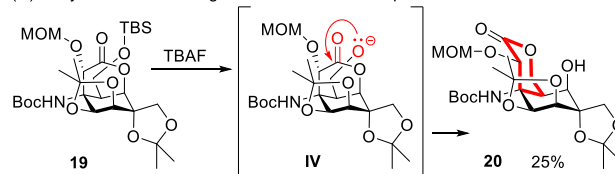
Enol ether **29**, which increased a methylene unit in the side chain, was synthesized from the known sulfone **23**^{7c} and pyrrolidinone **8** (Scheme 5). Compound **29** was

Scheme 4. (A) Synthetic Study on Producing Target Analogue **2** from Enol Ether **15**, (B) Recyclization According to Xu and Ciufolini's Report, and (C) Our New Synthetic Plan

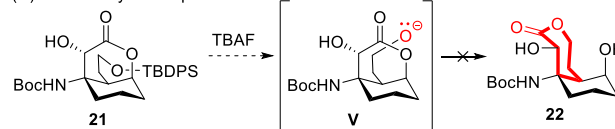
(A) Examination for the synthesis of **2** from enol ether **15** and the dead end



(B) Recyclization according to Xu and Ciufolini's report

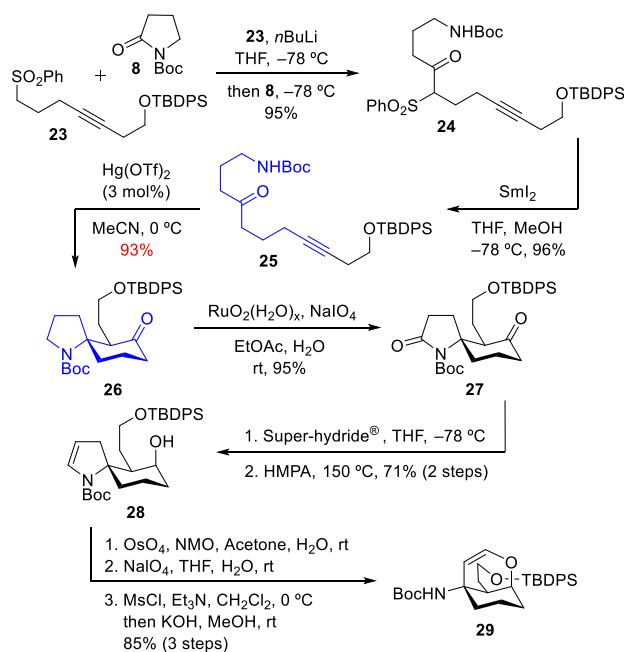


(C) Our new synthetic plan



prepared according to the same pathway as shown in Scheme 3. As the key reaction, the cycloisomerization reaction⁷ of ynone **25** yielded the desired spirocycle **26** in high yield (93%) and in a diastereoselective manner.

Scheme 5. Synthesis of Enol Ether **29**.



We tried to synthesize analogue **2** from enol ether **29** as shown in Scheme 6. In the next step, dihydroxylation of **29** with OsO₄ produced diol **30** as a single diastereomer²⁰ before the selective TEMPO oxidation¹⁸ of **30** provided lactone **21** (90%), along with diketone as a minor byproduct (9%). As expected, the desired alcohol **31** was obtained in high yield (98%) through the deprotection of a TBDPS group utilizing TBAF at 0 °C,²¹ while the unwanted seven-membered lactone **22** was not detected. Compound **31** was transformed to acetal **32** via the following sequence: 1) Nishizawa–Grieco elimination²² to construct a vinylic group, 2) ozonolysis of a vinylic group, and 3) acetal formation using PPTS and MeC(OMe)₃. Following removal of a Boc group with TMSOTf and 2,6-lutidine,⁶⁰ guanidinylation of the resultant amine produced **33** as a mixture of diastereomers in high yield (97%). Proceeding with reference to Fukuyama's report,^{6j} the deprotection under acidic conditions led to a mixture of mono-Cbz-protected precursor **34** and its anhydro form **35**. Finally, we managed to synthesize our target analogue, 11-*nor*-6,7,8-trideoxyTTX acetate (**2**·HOAc), through hydrogenolysis of a Cbz group and HPLC purification.⁵ⁿ An anhydro form of **2**·HOAc, 11-*nor*-4,9-anhydro-6,7,8-trideoxyTTX acetate (**36**·HOAc), was also obtained from **35** via the same process.

In summary, we have achieved the syntheses of 11-*nor*-6,7,8-trideoxyTTX, an unnatural tetrodotoxin analogue, and its anhydro form, 11-*nor*-4,9-anhydro-6,7,8-trideoxyTTX,²³ from a spirocyclic compound constructed through our Hg(OTf)₂-catalyzed cycloisomerization reaction of a linear substrate. The success of these syntheses encourages our new approach to TTX shown in Scheme 1B. The application of our synthetic strategy to natural TTX and other deoxy analogues is currently underway in our laboratory.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI. All experimental procedures, the spectroscopic data, and copies of ¹H- and ¹³C-NMR spectra.

AUTHOR INFORMATION

Corresponding Authors

Keisuke Nishikawa – Department of Chemistry, Graduate School of Science, Osaka City University, Sumiyoshi-ku, Osaka 558-8585, Japan; orcid.org/0000-0002-3170-8023; Email: knishi@sci.osaka-cu.ac.jp

Yoshiki Morimoto – Department of Chemistry, Graduate School of Science, Osaka City University, Sumiyoshi-ku, Osaka 558-8585, Japan; orcid.org/0000-0002-4770-3091; Email: morimoto@sci.osaka-cu.ac.jp

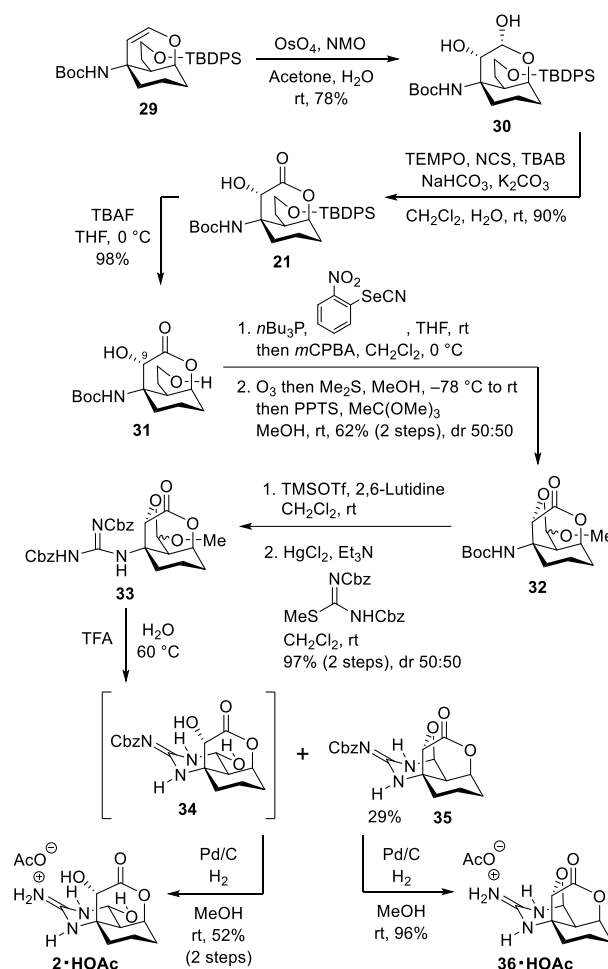
Authors

Takayuki Noguchi – Department of Chemistry, Graduate School of Science, Osaka City University, Sumiyoshi-ku, Osaka 558-8585, Japan

Seiho Kikuchi – Department of Chemistry, Graduate School of Science, Osaka City University, Sumiyoshi-ku, Osaka 558-8585, Japan

Takahiro Maruyama – Department of Chemistry, Graduate School of Science, Osaka City University, Sumiyoshi-ku, Osaka 558-8585, Japan

Scheme 6. Synthesis of 11-*nor*-6,7,8-TrideoxyTTX Acetate (**2**·HOAc) and Anhydro Form **36**·HOAc



Yusuke Araki – Department of Chemistry, Graduate School of Science, Osaka City University, Sumiyoshi-ku, Osaka 558-8585, Japan

Mari Yotsu-Yamashita – Graduate School of Agricultural Science, Tohoku University, Sendai, Miyagi 980-8572, Japan; orcid.org/0000-0002-5009-0409

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENT

This work was financially supported by the Astellas Foundation, the JGC-S Scholarship Foundation (Nikki Saneyoshi Scholarship), and JSPS KAKENHI Grant Number JP19K05461.

REFERENCES

- (1) Tahara, Y. Studies on the Puffer Fish Toxin. *J. Pharm. Soc. Jpn.* **1909**, *29*, 587–625.
- (2) (a) Goto, T.; Kishi, Y.; Takahashi, S.; Hirata, Y. Tetrodotoxin. *Tetrahedron* **1965**, *21*, 2059–2088. (b) Tsuda, K.; Ikuma, S.; Kawamura, M.; Tachikawa, R.; Sakai, K.; Tamura, C.; Amakasu, O. Tetrodotoxin. VII. On the Structures of Tetrodotoxin and Its Derivatives. *Chem. Pharm. Bull.* **1964**, *12*, 1357–1374. (c) Woodward, R. B. The Structure of Tetrodotoxin. *Pure Appl. Chem.* **1964**, *9*, 49–74.
- (3) Furusaki, A.; Tomiie, Y.; Nitta, I. The Crystal and Molecular Structure of Bromoanhydrotetrodoic Lactone Hydrobromide. *Bull. Chem. Soc. Jpn.* **1970**, *43*, 3325–3331.

- (4) (a) Narahashi, T.; Deguchi, T.; Urakawa, N.; Ohkubo, Y. Stabilization and Rectification of Muscle Fiber Membrane by Tetrodotoxin. *Am. J. Physiol.* **1960**, *198*, 934–938. (b) Narahashi, T.; Moore, J. W.; Scott, W. R. Tetrodotoxin Blockage of Sodium Conductance Increase in Lobster Giant Axons. *J. Gen. Physiol.* **1964**, *47*, 965–973. (c) Yasumoto, T.; Nagai, H.; Yasumura, D.; Michishita, T.; Endo, A.; Yotsu, M.; Kotaki, Y. Interspecies Distribution and Possible Origin of Tetrodotoxin. *Ann. N. Y. Acad. Sci.* **1986**, *479*, 44–51. (d) Terlau, H.; Heinemann, S. H.; Stühmer, W.; Pusch, M.; Conti, F.; Imoto, K.; Numa, S. Mapping the Site of Block by Tetrodotoxin and Saxitoxin of Sodium Channel II. *FEBS Lett.* **1991**, *293*, 93–96. (e) Rosker, C.; Lohberger, B.; Hofer, D.; Steinecker, B.; Quasthoff, S.; Schreibmayer, W. The TTX Metabolite 4,9-Anhydro-TTX is a Highly Specific Blocker of the $\text{Na}_{v1.6}$ Voltage-Dependent Sodium Channel. *Am. J. Physiol. Cell Physiol.* **2007**, *293*, C783–C789. (f) Tsukamoto, T.; Chiba, Y.; Wakamori, M.; Yamada, T.; Tsunogae, S.; Cho, Y.; Sakakibara, R.; Imazu, T.; Tokoro, S.; Satake, Y.; Adachi, M.; Nishikawa, T.; Yotsu-Yamashita, M.; Konoki, K. Differential Binding of Tetrodotoxin and Its Derivatives to Voltage-Sensitive Sodium Channel Subtypes ($\text{Na}_{v1.1}$ to $\text{Na}_{v1.7}$). *Br. J. Pharmacol.* **2017**, *174*, 3881–3892. (g) Shen, H.; Li, Z.; Jiang, Y.; Pan, X.; Wu, J.; Cristofori-Armstrong, B.; Smith, J. J.; Chin, Y. K. Y.; Lei, J.; Zhou, Q.; King, G. F.; Yan, N. Structural Basis for the Modulation of Voltage-Gated Sodium Channels by Animal Toxins. *Science* **2018**, *362*, eaau2596. (h) Shen, H.; Liu, D.; Wu, K.; Lei, J.; Yan, N. Structures of Human $\text{Na}_{v1.7}$ Channel in Complex with Auxiliary Subunits and Animal Toxins. *Science* **2019**, *363*, 1303–1308.
- (5) (a) For a review, see: Bane, V.; Lehane, M.; Dikshit, M.; O'Riordan, A.; Furey, A. Tetrodotoxin: Chemistry, Toxicity, Source, Distribution and Detection. *Toxins* **2014**, *6*, 693–755. (b) Nakamura, M.; Yasumoto, T. Tetrodotoxin Derivatives in Puffer Fish. *Toxicon* **1985**, *23*, 271–276. (c) Yasumoto, T.; Yotsu, M.; Murata, M.; Naoki, H. New Tetrodotoxin Analogs from the Newt *Cynops ensicauda*. *J. Am. Chem. Soc.* **1988**, *110*, 2344–2345. (d) Endo, A.; Khora, S. S.; Murata, M.; Naoki, H.; Yasumoto, T. Isolation of 11-norTetrodotoxin-6(R)-ol and Other Tetrodotoxin Derivatives from the Puffer *Fugu Niphobles*. *Tetrahedron Lett.* **1988**, *29*, 4127–4128. (e) Khora, S. S.; Yasumoto, T. Isolation of 11-oxoTetrodotoxin from the Puffer *Arothron nigropunctatus*. *Tetrahedron Lett.* **1989**, *30*, 4393–4394. (f) Yotsu, M.; Hayashi, Y.; Kohra, S. S.; Sato, S.; Yasumoto, T. Isolation and Structural Assignment of 11-norTetrodotoxin-6(S)-ol from the Puffer *Arothron nigropunctatus*. *Biosci. Biotechnol. Biochem.* **1992**, *56*, 370–371. (g) Yotsu-Yamashita, M.; Sugimoto, A.; Takai, A.; Yasumoto, T. Effects of Specific Modifications of Several Hydroxyls of Tetrodotoxin on Its Affinity to Rat Brain Membrane. *J. Pharmacol. Exp. Ther.* **1999**, *289*, 1688–1696. (h) Jang, J.-H.; Yotsu-Yamashita, M.; 6,11-Dideoxytetrodotoxin from the Puffer Fish, *Fugu pardalis*. *Toxicon* **2007**, *50*, 947–951. (i) Yotsu-Yamashita, M.; Abe, Y.; Kudo, Y.; Ritson-Williams, R.; Paul, V. J.; Konoki, K.; Cho, Y.; Adachi, M.; Imazu, T.; Nishikawa, T.; Isobe, M. First Identification of 5,11-Dideoxytetrodotoxin in Marine Animals, and Characterization of Major Fragment Ions of Tetrodotoxin and Its Analogs by High Resolution ESI-MS/MS. *Mar. Drugs* **2013**, *11*, 2799–2813. (j) Kudo, Y.; Finn, J.; Fukushima, K.; Sakugawa, S.; Cho, Y.; Konoki, K.; Yotsu-Yamashita, M. Isolation of 6-Deoxytetrodotoxin from the Pufferfish, *Takifugu pardalis*, and a Comparison of the Effects of the C-6 and C-11 Hydroxy Groups of Tetrodotoxin on Its Activity. *J. Nat. Prod.* **2014**, *77*, 1000–1004. (k) Kudo, Y.; Yamashita, Y.; Mebs, D.; Cho, Y.; Konoki, K.; Yasumoto, T.; Yotsu-Yamashita, M. C5-C10 Directly Bonded Tetrodotoxin Analogues: Possible Biosynthetic Precursors of Tetrodotoxin from Newts. *Angew. Chem., Int. Ed.* **2014**, *53*, 14546–14549. (l) Puiling, C. G.; Kudo, Y.; Cho, Y.; Konoki, K.; Yotsu-Yamashita, M. Tetrodotoxin and Its Analogues in the Pufferfish *Arothron hispidus* and *A. nigropunctatus* from the Solomon Islands: A Comparison of Their Toxin Profiles with the Same Species from Okinawa, Japan. *Toxins* **2015**, *7*, 3436–3454. (m) Kudo, Y.; Yasumoto, T.; Mebs, D.; Cho, Y.; Konoki, K.; Yotsu-Yamashita, M. Cyclic Guanidine Compounds from Toxic Newts Support the Hypothesis that Tetrodotoxin is Derived from a Monoterpene. *Angew. Chem., Int. Ed.* **2016**, *55*, 8728–8731. (n) Ueyama, N.; Sugimoto, K.; Kudo, Y.; Onodera, K.-I.; Cho, Y.; Konoki, K.; Nishikawa, T.; Yotsu-Yamashita, M. Spiro Bicyclic Guanidino Compounds from Pufferfish: Possible Biosynthetic Intermediates of Tetrodotoxin in Marine Environments. *Chem. - Eur. J.* **2018**, *24*, 7250–7258. (o) Kudo, Y.; Yotsu-Yamashita, M. Isolation and Biological Activity of 8-*epi*Tetrodotoxin and the Structure of a Possible Biosynthetic Shunt Product of Tetrodotoxin, Cep-226A, from the Newt *Cynops ensicauda popei*. *J. Nat. Prod.* **2019**, *82*, 1656–1663. (p) Kudo, Y.; Hanifin, C.; Kotaki, Y.; Yotsu-Yamashita, M. Structures of N-Hydroxy-Type Tetrodotoxin Analogues and Bicyclic Guanidinium Compounds Found in Toxic Newts. *J. Nat. Prod.* **2020**, *83*, 2706–2717.
- (6) For a review, see: (a) Makarova, M.; Rycek, L.; Hajicek, J.; Baidilov, D.; Hudlicky, T. Tetrodotoxin: History, Biology, and Synthesis. *Angew. Chem., Int. Ed.* **2019**, *58*, 18338–18387. For racemic synthesis, see: (b) Kishi, Y.; Aratani, M.; Fukuyama, T.; Nakatsubo, F.; Goto, T.; Inoue, S.; Tanino, H.; Sugiura, S.; Kakoi, H. Synthetic Studies on Tetrodotoxin and Related Compounds. III. Stereospecific Synthesis of an Equivalent of Acetylated Tetradamine. *J. Am. Chem. Soc.* **1972**, *94*, 9217–9219. (c) Kishi, Y.; Fukuyama, T.; Aratani, M.; Nakatsubo, F.; Goto, T.; Inoue, S.; Tanino, H.; Sugiura, S.; Kakoi, H. Synthetic Studies on Tetrodotoxin and Related Compounds. IV. Stereospecific Total Syntheses of DL-Tetrodotoxin. *J. Am. Chem. Soc.* **1972**, *94*, 9219–9221. (d) Sato, K.; Akai, S.; Sugita, N.; Ohsawa, T.; Kogure, T.; Shoji, H.; Yoshimura, J. Novel and Stereoccontrolled Synthesis of (\pm)-Tetrodotoxin from *myo*-Inositol. *J. Org. Chem.* **2005**, *70*, 7496–7504. For asymmetric synthesis, see: (e) Ohyabu, N.; Nishikawa, T.; Isobe, M. First Asymmetric Total Synthesis of Tetrodotoxin. *J. Am. Chem. Soc.* **2003**, *125*, 8798–8805. (f) Hinman, A.; Du Bois, J. A Stereoselective Synthesis of (–)-Tetrodotoxin. *J. Am. Chem. Soc.* **2003**, *125*, 11510–11511. (g) Nishikawa, T.; Urabe, D.; Isobe, M. An Efficient Total Synthesis of Optically Active Tetrodotoxin. *Angew. Chem., Int. Ed.* **2004**, *43*, 4782–4785. (h) Sato, K.; Akai, S.; Shoji, H.; Sugita, N.; Yoshida, S.; Nagai, Y.; Suzuki, K.; Nakamura, Y.; Kajihara, Y.; Funabashi, M.; Yoshimura, J. Stereoselective and Efficient Total Synthesis of Optically Active Tetrodotoxin from D-Glucose. *J. Org. Chem.* **2008**, *73*, 1234–1242. (i) Akai, S.; Seki, H.; Sugita, N.; Kogure, T.; Nishizawa, N.; Suzuki, K.; Nakamura, Y.; Kajihara, Y.; Yoshimura, J.; Sato, K. Total Synthesis of (–)-Tetrodotoxin from D-Glucose: A New Route to Multi-Functionalized Cyclitol Employing the Ferrier(II) Reaction toward (–)-Tetrodotoxin. *Bull. Chem. Soc. Jpn.* **2010**, *83*, 279–287. (j) Maehara, T.; Motoyama, K.; Toma, T.; Yokoshima, S.; Fukuyama, T. Total Synthesis of (–)-Tetrodotoxin and 11-norTTX-6(R)-ol. *Angew. Chem., Int. Ed.* **2017**, *56*, 1549–1552. (k) Murakami, K.; Toma, T.; Fukuyama, T.; Yokoshima, S. Total Synthesis of Tetrodotoxin. *Angew. Chem., Int. Ed.* **2020**, *59*, 6253–6257. For synthesis of analogues, see: (l) Nishikawa, T.; Urabe, D.; Yoshida, K.; Iwabuchi, T.; Asai, M.; Isobe, M. Stereocontrolled Synthesis of 8,11-Dideoxytetrodotoxin, Unnatural Analogue of Puffer Fish Toxin. *Org. Lett.* **2002**, *4*, 2679–2682. (m) Nishikawa, T.; Urabe, D.; Yoshida, K.; Iwabuchi, T.; Asai, M.; Isobe, M. Total Syntheses of 11-Deoxytetrodotoxin and 8,11-Dideoxytetrodotoxin. *Pure Appl. Chem.* **2003**, *75*, 251–257. (n) Nishikawa, T.; Urabe, D.; Yoshida, K.; Iwabuchi, T.; Asai, M.; Isobe, M. Stereocontrolled Synthesis of 8,11-Dideoxytetrodotoxin, An Unnatural Analogue of Puffer Fish Toxin. *Chem. - Eur. J.* **2004**, *10*, 452–462. (o) Umezawa, T.; Hayashi, T.; Sakai, H.; Teramoto, H.; Yoshikawa, T.; Izumida, M.; Tamatani, Y.; Hirose, T.; Ohfune, Y.; Shinada, T. Total Synthesis of (–)-5,6,11-Trideoxytetrodotoxin and Its 4-Epimer. *Org. Lett.* **2006**, *8*, 4971–4974. (p) Adachi, M.; Imazu, T.; Isobe, M.; Nishikawa, T. *J. Org. Chem.* **2013**, *78*, 1699–1705. (q) Satake, Y.; Adachi, M.; Tokoro, S.; Yotsu-Yamashita, M.; Isobe, M.; Nishikawa, T. Synthesis of 5- and 8-Deoxytetrodotoxin. *Chem. - Asian J.* **2014**, *9*, 1922–1932. (r) Adachi, M.; Sakakibara, R.; Satake, Y.; Isobe, M.; Nishikawa, T. Synthesis of 5,6,11-Trideoxytetrodotoxin. *Chem. Lett.* **2014**, *43*, 1719–1721. (s) Adachi, M.; Miyasaka, T.; Kudo, Y.; Sugimoto, K.; Yotsu-Yamashita, M.; Nishikawa, T. Total Syntheses and Determination of Absolute Configurations of Cep-

212 and Cep-210, Predicted Biosynthetic Intermediates of Tetrodotoxin Isolated from Toxic Newt. *Org. Lett.* **2019**, *21*, 780–784.

(7) (a) Nishikawa, K.; Kikuchi, S.; Ezaki, S.; Koyama, T.; Nokubo, H.; Kodama, T.; Tachi, Y.; Morimoto, Y. Total Synthesis of (–)-Lepadiformine A Utilizing Hg(OTf)₂-Catalyzed Cycloisomerization Reaction. *Org. Lett.* **2015**, *17*, 5772–5775. (b) Nishikawa, K.; Yamauchi, K.; Kikuchi, S.; Ezaki, S.; Koyama, T.; Nokubo, H.; Matsumura, K.; Kodama, T.; Kumagai, M.; Morimoto, Y. Total Syntheses of Lepadiformine Marine Alkaloids with Enantiodivergency, Utilizing Hg(OTf)₂-Catalyzed Cycloisomerization Reaction and Their Cytotoxic Activities. *Chem. - Eur. J.* **2017**, *23*, 9535–9545. (c) Matsumura, K.; Nishikawa, K.; Yoshida, H.; Doe, M.; Morimoto, Y. Formal Total Synthesis of Histrionicotoxin Alkaloids via Hg(OTf)₂-Catalyzed Cycloisomerization and SmI₂-Induced Ring Expansion. *RSC Adv.* **2018**, *8*, 11296–11303.

(8) Choi, S.-R.; Breugst, M.; Houk, K. N.; Poulter, C. D. δ -Deuterium Isotope Effects as Probes for Transition-State Structures of Isoprenoid Substrates. *J. Org. Chem.* **2014**, *79*, 3572–3580.

(9) Giovannini, A.; Savoia, D.; Umani-Ronchi, A. Organometallic Ring-Opening Reactions of *N*-Acyl and *N*-Alkoxy carbonyl Lactams. Synthesis of Cyclic Imines. *J. Org. Chem.* **1989**, *54*, 228–234.

(10) Molander, G. A.; Hahn, G. Lanthanides in Organic Synthesis. 2. Reduction of α -Heterosubstituted Ketones. *J. Org. Chem.* **1986**, *51*, 1135–1138.

(11) The relative configurations of compounds **6**, **14**, **16–18**, **26**, **28**, **30**, **2•HOAc**, and **36•HOAc** were unambiguously determined by their NOESY spectra; see the [Supporting Information \(SI\)](#).

(12) The Ph₃PAuOTf-catalyzed cycloisomerization reaction of **7** afforded the desired **6** only in low yield (<25%).

(13) For a discussion about the more detailed transition states, see ref. (7).

(14) Shinada, T.; Yoshida, H.; Ohfuné, Y. Efficient Synthesis of (R)- and (S)- α -(Hydroxymethyl)pyroglutamic Acid Esters from L-Proline. *Synthesis* **2009**, 3751–3756.

(15) (a) Monson, R. S. Solvent Catalyzed Dehydration of Alcohols in Hexamethylphosphoric Triamide. *Tetrahedron Lett.* **1971**, *7*, 567–570. (b) Dieter, R. K.; Sharma, R. R. A Facile Preparation of Enecarbamates. *J. Org. Chem.* **1996**, *61*, 4180–4184.

(16) Parida, K. N.; Moorthy, J. N. Oxidation Cascade with Oxone: Cleavage of Olefins to Carboxylic Acids. *Tetrahedron* **2014**, *70*, 2280–2285.

(17) The OsO₄-catalyzed dihydroxylation of **15** resulted in the recovery of the starting material.

(18) Einhorn, J.; Einhorn, C.; Ratajczak, F.; Pierre, J.-L. Efficient and Highly Selective Oxidation of Primary Alcohols to Aldehydes by *N*-Chlorosuccinimide Mediated by Oxoammonium Salts. *J. Org. Chem.* **1996**, *61*, 7452–7454.

(19) Xu, S.; Ciufolini, M. A. Formal Synthesis of (±)-Tetrodotoxin via the Oxidative Amidation of a Phenol: On the Structure of the Sato Lactone. *Org. Lett.* **2015**, *17*, 2424–2427.

(20) Dihydroxylation of **29** using Oxone® and NaHCO₃ gave the desired **30** in 64% yield, along with lactol which would oxidatively be cleaved from **30** (25%).

(21) The TBDPS deprotection at room temperature afforded a mixture of two diastereomers, diol **31** and its 9-epimer (dr 59:41), see the [SI](#). We selectively synthesized 9-epimer (dr 90:10) through the NaBH₄ reduction of diketone which produced as a byproduct in the oxidation of **30**, followed by the deprotection of a TBDPS protecting group, to obtain the spectral data of 9-epimer. The stereochemistry of 9-epimer was clearly assigned based on the NOESY spectrum; see the [SI](#).

(22) Grieco, P. A.; Gilman, S.; Nishizawa, M. Organoselenium Chemistry. A Facile One-Step Synthesis of Alkyl Aryl Selenides from Alcohols. *J. Org. Chem.* **1976**, *41*, 1485–1486.

(23) With synthetic 11-*nor*-6,7,8-trideoxyTTX acetate (**2•HOAc**) and an anhydro form **36•HOAc** in hand, we evaluated their effects on cell viability in the presence of ouabain and veratridine in mouse neuroblastoma Neuro-2a cells. The inhibition activity of voltage-gated sodium channels by TTX analogues was estimated from their reduction of the cytotoxicity of ouabain and veratridine (See: (a) Kogure, K.; Tamplin, M. L.; Simidu, U.; Colwell, R. R. A Tissue Culture Assay for Tetrodotoxin, Saxitoxin and Related Toxins. *Toxicon* **1988**, *26*, 191–197. (b) Yotsu-Yamashita, M.; Urabe, D.; Asai, M.; Nishikawa, T.; Isobe, M. Biological Activity of 8,11-Dideoxytetrodotoxin: Lethality to Mice and the Inhibitory Activity to Cytotoxicity of Ouabain and Veratridine in Mouse Neuroblastoma Cells, Neuro-2a. *Toxicon* **2003**, *42*, 557–560.). Unfortunately, two analogues did not show any activity (all IC₅₀ values >100 μ M).